

# GMOMETHODS: EU DATABASE OF REFERENCE METHODS

## Qualitative PCR method for detection of nopaline synthase terminator

*Element specific*

Last updated 24/5/2017

### 1. GENERAL INFORMATION

<b>Target genetic element</b>	Nopaline synthase terminator (T-nos) from <i>Agrobacterium tumefaciens</i>
<b>PCR Assay</b>	Simplex Real Time
<b>Detection</b>	TaqMan
<b>Compendium Reference</b>	QL-ELE-00-011

### 2. VALIDATION DATA

<b>Collaborative trial coordinator</b>	Federal Office of Consumer Protection and Food Safety (BVL)
<b>Test material applied in collaborative trial</b>	DNA
<b>Materials used for calibration/controls</b>	CRM BF-415b and BF-415c (JRC-IRMM)

#### Tested GM Events

Event Name	Unique Identifier	Crop Name
NK603	MON-00603-6	Zea mays

#### Collaborative Trial Description

Each participant received 18 blind DNA samples of which 12 were prepared from GMO CRM (0.1% or 0.5% maize event NK603) and six from non-GM maize flour. In addition, participants received a positive DNA target control consisting of a solution of 0.5% mass fraction event NK603 DNA and solutions of primers, probe and a commercial reagent kit specific for the laboratory PCR platform. 12 laboratories used real-time PCR equipment designed for plastic reaction vials while other 12 laboratories equipped with a LightCycler instrument used glass capillaries. The participants were requested to submit the cycle threshold (Ct) values obtained in the T-nos real-time PCR with each sample and to report whether the sample was considered positive or negative.

#### Method Performance

<b>LOD Relative</b>	≤0.1%	<b>LOD Absolute</b>	8 target copies
---------------------	-------	---------------------	-----------------

**Values determined in the collaborative trial**

<b>False positive (%)</b>	2.2%
<b>False negative (%)</b>	0%

	<b>Test Level (%)</b>		
	0	0.1	0.5
<b>Specificity (%)</b>	98%		
<b>Sensitivity (%)</b>		100%	100%

Unit of Measurement Test Level % GMO (w/w)

**Comment**

The performance values reported in the table for the 0.1% and 0.5% (w/w) levels were obtained with PCR equipment designed for plastic reaction vials. The false positives rate instead combines the data from both real-time PCR thermocyclers. The method was also positively tested in-house on samples of other T-nos containing GM-events such as soybean GTS-40-3-2, maize Bt11, NK603, GA21, MON809, MON863, CBH-351, MIR604, papaya SunUp, tomato Zeneca, canola MS1/RF1, MS8/RF3, and rice Oxy235 and BT63.

**3. REFERENCES**

Reiting R, Broll H, Waiblinger H-U, Grohmann L. Collaborative Study of a T-nos Real-Time PCR Method for Screening of Genetically Modified Organisms in Food Products. *J. Verbr. Lebensm.* 2007; 2: 116-121

DOI 10.1007/s00003-007-0189-4

ISO 21569:2005/Amd 1:2013: Foodstuffs--Methods of analysis for the detection of genetically modified organisms and derived products -- Qualitative nucleic acid based methods Amendment 1

**4. PRIMERS AND PROBES SEQUENCES**

**GM-target(s)** Nopaline synthase terminator (T-nos) from *Agrobacterium tumefaciens*

<b>Primer Forward</b>	5'-CATGTAATGCATGACGTTATTTATG-3'
<b>Target element</b>	T-nos
<b>Primer Reverse</b>	5'-TTGTTTTCTATCGCGTATTAAATGT-3'
<b>Target element</b>	T-nos
<b>Amplicon length</b>	84 bp
<b>Probe</b>	5'-FAM-ATGGGTTTTTATGATTAGAGTCCCGCAA-TAMRA-3'



## 5. PCR REACTIONS SETUP

### GM-target(s)

Reagent	Final Concentration
QuantiTect Probe PCR Kit (Qiagen) 2x	1x
Primer Fw	0.40 $\mu\text{mol/L}$
Primer Rev	0.40 $\mu\text{mol/L}$
Probe	0.10 $\mu\text{mol/L}$
Template DNA	100 ng
Final Volume	25 $\mu\text{L}$

## 6. AMPLIFICATION CONDITIONS

### GM-target(s)

Stage	Temperature	Time	NoCycles
Activation/Initial Denaturation	95°C	900"	1
Denaturation	94°C	15"	
Annealing & Extension	60°C	60"	
Denaturing, Annealing & Extension			45